Acta Crystallographica Section C Crystal Structure Communications

ISSN 0108-2701

# O-Benzyl-*N-tert*-butoxycarbonyl-*N*-methyl-L-tyrosine

# Elżbieta Jankowska,<sup>a</sup> Mirosław Gilski,<sup>b</sup>\* Mariusz Jaskólski,<sup>c</sup> Zbigniew Grzonka<sup>a</sup> and Leszek Łankiewicz<sup>a</sup>

<sup>a</sup>Faculty of Chemistry, University of Gdańsk, Sobieskiego 18, 80-952 Gdańsk, Poland, <sup>b</sup>Department of Crystallography, Faculty of Chemistry, A. Mickiewicz University, Grunwaldzka 6, 60-780 Poznań, Poland, and <sup>c</sup>Center for Biocrystallographic Research, Institute of Bioorganic Chemistry, Polish Academy of Sciences, Noskowskiego 12/14, 61-704 Poznań, Poland Correspondence e-mail: mirek@amu.edu.pl

Received 27 February 2002 Accepted 18 April 2002 Online 31 May 2002

The crystal structure of the title compound, alternatively called 3-[4-(benzyloxy)phenyl]-2-(*N*-tert-butoxycarbonyl-*N*-methylamino)propionic acid,  $C_{22}H_{27}NO_5$ , has been studied in order to examine the role of *N*-methylation as a determinant of peptide conformation. The conformation of the *tert*-butoxycarbonyl group is *trans*-*trans*. The side chain has a folded conformation and the two phenyl rings are effectively perpendicular to one another. The carboxylate hydroxyl group and the urethane carbonyl group form a strong intermolecular O-H···O hydrogen bond.

## Comment

Conformational studies of peptides and pseudopeptides (peptidomimetics) represent an ongoing project in our laboratory aimed at finding new biologically active peptide analogues and new protease inhibitors. *N*-Alkylated amino acids occur in many natural compounds, such as antibiotics or toxins, and are widely used in the synthesis of peptide analogues. As an example, incorporation of *N*-methylglycine (sarcosine) and *N*-methylalanine at position 7 of oxytocin has led to new derivatives with interesting properties (Grzonka *et al.*, 1983; Gazis *et al.*, 1984). The well known and widely



used immunosuppressant drug cyclosporin A contains seven *N*-methylated amino acid residues (Loosli *et al.*, 1985). The *N*-alkylated peptide bond is more resistant to enzymatic cleavage. The *cis*-*trans* isomerization of *N*-alkylated peptide



Figure 1

Displacement-ellipsoid view (50% probability level) of the title compound with the atom-numbering scheme.

bonds is shifted towards the *cis* isomer. Finally, incorporation of *N*-alkylamino acid residues into the polypeptide chain decreases the possibility of hydrogen-bond formation, and at the same time, increases the hydrophobicity of such peptide fragments (Spatola, 1983). The present study is a contribution to our understanding of the structural role of *N*-methylation in amino acids and peptide derivatives.

The molecular structure and atom-numbering scheme of the title compound, (I), is shown in Fig. 1. A selection of bond distances and angles is given in Table 1 together with some notable torsion angles. The bond lengths within the *tert*-butoxycarbonyl group are very similar (except for the C19– O4 bond, which is lengthened by nearly 0.02 Å) to the average lengths for this blocking group compiled by Benedetti *et al.* (1980). There is one notable variation from these averages in the bond angles involving the urethane bond. Thus, the C18– O4–C19 angle [123.1 (2)°] is 2° wider than the computed averages for these urethane linkages (Table 1). The conformation of the *tert*-butoxycarbonyl group, characterized by the torsion angles  $\Theta_0$  (C19–O4–C18–O3) and  $\omega_0$  (O4–C18–N1–C2), is *trans-trans* (Table 1). This is opposite to most of the published X-ray structures of compounds containing the



**Figure 2** The crystal packing of the title compound viewed along *c*.

*tert*-butoxycarbonyl group at a tertiary N atom (e.g. proline or sarcosine derivatives), which usually have a cis-urethane bond (Benedetti et al., 1980). The deviation from planarity of the urethane bond is in a typical range found for tert-butoxycarbonylamino acid and peptide derivatives (Sopková et al., 1996; Banumathi et al., 1999). The backbone conformation is characterized by the torsion angles  $\Phi$  (C18–N1–C2–C1) and  $\Psi$  (O1-C1-C2-N1), and the conformation of the side chain is characterized by  $\chi_1$  (N1-C2-C3-C4) and  $\chi_2$  (C2-C3-C4-C5) (Table 1). Therefore, the side chain has a folded conformation. The O5-C10 bond is roughly coplanar with the tyrosine ring  $[C6-C7-O5-C10 = 1.3 (4)^{\circ}]$ , but is perpendicular to the benzyl ring  $[O5-C10-C11-C12 = -85.7 (3)^{\circ}]$ . With the O5-C10 bond in the trans orientation [C7-O5- $C10-C11 = -176.9 (2)^{\circ}$ ], the two phenyl rings are thus perpendicular to one another [interplanar angle =  $86.5 (5)^{\circ}$ ]. The tyrosine and benzyl rings are planar, with  $\chi^2$  values of 43.1 and 11.1°, respectively. In the crystal packing, the tyrosine and benzyl moieties form layers, with the *tert*-butoxycarbonyl groups located between them (Fig. 2).

The carboxylate hydroxyl and the urethane carbonyl group form a strong intermolecular O-H···O hydrogen bond  $[O1 \cdots O3^{i} 2.679 (2) \text{ Å and } O1 - H1 \cdots O3^{i} 170 (3)^{\circ}; \text{ symmetry}$ code: (i) -x - 1,  $y - \frac{1}{2}$ ,  $-z - \frac{1}{2}$ ] (Fig. 2). A similar hydrogen bond was also reported in the crystal structure of tertbutoxycarbonylphenylalanine (Bats et al., 1980).

## Experimental

The title compound was synthesized from O-benzyl-tert-butoxycarbonyl-L-tyrosine and methyl iodide in the presence of KH and crown ether (18-C-6) (Schuman et al., 1983). The tyrosine derivative thus formed is suitable for peptide synthesis both in solution and in the solid phase via peptide-chain elongation because it has a protected amino group (Boc) and an unprotected carboxyl group. Single crystals of (I) (m.p. = 400-401 K) were obtained by slow evaporation from an ethyl acetate/petroleum ether solution.

## Crystal data

C <sub>22</sub> H <sub>27</sub> NO <sub>5</sub>	Mo $K\alpha$ radiation
$M_r = 385.45$	Cell parameters from 6728
Orthorhombic, $P2_12_12_1$	reflections
a = 9.346 (2) Å	$\theta = 3.4-29.6^{\circ}$
b = 11.413 (2) Å	$\mu = 0.09 \text{ mm}^{-1}$
c = 19.493 (4)  Å	T = 100 (2)  K
V = 2079.2 (7) Å <sup>3</sup>	Prism, colourless
Z = 4	$0.35 \times 0.30 \times 0.25 \text{ mm}$
$D_{\rm m} = 1.231 {\rm Mg}{\rm m}^{-3}$	

#### Table 1

Selected geometric parameters  $(\hat{A}, \circ)$ .

C1-O2	1.201 (3)	C17-N1	1.459 (3)
C1-O1	1.327 (3)	C18-O3	1.229 (3)
C2-N1	1.462 (3)	C18-O4	1.340 (3)
C7-O5	1.367 (3)	C18-N1	1.356 (3)
C10-O5	1.440 (3)	C19-O4	1.491 (3)
$\begin{array}{c} 01 - C1 - C2 - N1 \\ N1 - C2 - C3 - C4 \\ C2 - C3 - C4 - C5 \\ O5 - C10 - C11 - C12 \\ C6 - C7 - O5 - C10 \end{array}$	175.9 (2) -54.3 (3) -48.6 (4) -85.7 (3) 1.3 (4)	C11-C10-O5-C7 O3-C18-O4-C19 O4-C18-N1-C2 C1-C2-N1-C18	-176.9 (2) 4.3 (4) 172.3 (2) -103.2 (3)

#### Data collection

Kuma KM-4 CCD diffractometer	$R_{\rm int} = 0.044$
w scans	$\theta_{\rm max} = 29.6^{\circ}$
11 736 measured reflections	$h = -12 \rightarrow 9$
3067 independent reflections	$k = -15 \rightarrow 15$
2602 reflections with $I > 2\sigma(I)$	$l = -26 \rightarrow 25$
D offer over over	

## Refinement

Refinement on $F^2$	$w = 1/[\sigma^2(F_o^2) + (0.0369P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.048$	+ 0.4413P]
$wR(F^2) = 0.112$	where $P = (F_o^2 + 2F_c^2)/3$
S = 1.34	$(\Delta/\sigma)_{\rm max} < 0.001$
3067 reflections	$\Delta \rho_{\rm max} = 0.36 \text{ e } \text{\AA}^{-3}$
257 parameters	$\Delta \rho_{\rm min} = -0.34 \text{ e } \text{\AA}^{-3}$
H atoms treated by a mixture of	
independent and constrained	
refinement	

The position and isotropic displacement parameter of the hydroxyl H atom were refined. All other H atoms were generated geometrically and treated as riding atoms (C-H = 0.93-0.98 Å). The Friedel-equivalent reflections were merged, as attempted refinement of the Flack (1983) parameter was inconclusive [refined value 0.5(10)]. The absolute configuration was set by reference to the known configuration of L-tyrosine.

Data collection: KM4CCD Software (Kuma, 1995-1999); cell refinement: KM4CCD Software; data reduction: KM4CCD Software; program(s) used to solve structure: SHELXS97 (Sheldrick, 1997); program(s) used to refine structure: SHELXL97 (Sheldrick, 1997); molecular graphics: Stereochemical Workstation Operation Manual (Siemens, 1989).

The present work was supported by the Polish Committee for Scientific Research (KBN) (grant No. 1245/T09/97/17).

Supplementary data for this paper are available from the IUCr electronic archives (Reference: GD1196). Services for accessing these data are described at the back of the journal.

### References

- Banumathi, S., Velmurugan, D., Subramanian, E., Ashish, N. & Kishore, R. (1999). Acta Cryst. C55, 78-79.
- Bats, J. W., Fuess, H., Kessler, H. & Schuck, R. (1980). Chem. Ber. 113, 520-530.
- Benedetti, E., Pedone, C., Toniolo, C., Nemethy, G., Pottle, M. S. & Scheraga, H. A. (1980). Int. J. Pept. Protein Res. 16, 165-172.
- Flack, H. D. (1983). Acta Cryst. A39, 876-881.
- Gazis, D., Schwartz, I. L., Lammek, B. & Grzonka, Z. (1984). Int. J. Pept. Protein Res. 23, 78-83.
- Grzonka, Z., Lammek, B., Kasprzykowski, F., Gazis, D. & Schwartz, I. L. (1983). J. Med. Chem. 26, 555-559.
- Kuma (1995-1999). KM4CCD Software. Version 1.161. Kuma Diffraction Instruments GmbH, Wrocław, Poland.
- Loosli, H.-R., Kessler, H., Oschkinat, H., Weber, H.-P., Petcher, T. J. & Widmer, A. (1985). Helv. Chim. Acta, 68, 682-704.
- Schuman, R. T., Gesellchen, P. D., Smithwick, E. L. & Frederickson, R. C. A. (1983). Peptides, Structure and Function, edited by V. J. Hruby & D. H. Rich, pp. 143-146. Rockford: Pierce Chemical Company.
- Sheldrick, G. M. (1997). SHELXL97 and SHELXS97. University of Göttingen, Germany.
- Siemens (1989). Stereochemical Workstation Operation Manual. Release 3.4. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.
- Sopková, J., Cisařová, I. & Arnold, A. (1996). Acta Cryst. C52, 2903–2905.
- Spatola, A. F. (1983). Chemistry and Biochemistry of Amino Acids, Peptides, and Proteins, edited by B. Weinstein, pp. 267-357. New York: Marcel Dekker Inc.